

Physiological Response and Tolerance of Genotypes of Shallots (*Allium ascalonicum* L.) under Shade Stress

Ni Ketut Sukasni¹, Taufik Fauzi¹, Farid Hemon¹

¹ *University of Mataram*

Jl. Majapahit No 62 Mataram, Nusa Tenggara Barat, Indonesia

DOI: [10.22178/pos.86-13](https://doi.org/10.22178/pos.86-13)

LCC Subject Category: S1-(972)

Received 28.09.2022

Accepted 26.10.2022

Published online 31.10.2022

Corresponding Author:

Ni Ketut Sukasni

sukasmi@unram.ac.id

© 2022 The Authors. This article is licensed under a Creative Commons Attribution 4.0

License 

Abstract. This study aims to determine several shallot genotypes' physiological responses and tolerance to shade stress. The experimental design used was a completely randomised design (CRD) consisting of 6 genotypes, namely: G1 (Keta Monca), G2 (Ampanan), G3 (Super Philip), G4 (Thailand Nganjuk), G5 (Bali Karet), G6 (Lokananta). Each treatment was repeated five times. Planting was done using polybags measuring 35x35 cm; each polybag planted two bulbs, 2-3 cm deep, with a spacing of 20x20 cm. Shade treatment used 65.00% parent with a shade height of 2 m, a width of 1.5 m, and a shade length of 3 m. The results of the physiological response research showed that the parameters of N-Total, protein, carbohydrates, and carotenoids showed significant differences in the genotype of shallots and indicated an interaction between treatments. The parameters of chlorophyll a, chlorophyll b, and proline showed no significant difference and no interaction between treatments. The genotypes of shallots sensitive to shade resistance were genotype Super Philip, Thailand Nganjuk, and Bali Karet. The moderate tolerant genotype was Ampanan, and the susceptible genotype was Keta Monca.

Keywords: shade; proline; chlorofill.

INTRODUCTION

Shallot (*Allium ascalonicum* L.) is one of the horticultural commodities that have been developed nationally. The prospect of onion cultivation is excellent because it can improve farmers' living standards. It has a high economic and industrial value and export potential [1].

Indonesia's centre of shallot production is Java Island, with a total output of 956,652 tons or about 77.53% of the total national shallot production. Central Java province is the largest shallot producer, producing 519,356 tons or 42.09% of the total national shallot production, followed by East Java and West Java. West Nusa Tenggara is the largest shallot-producing province outside Java, with a show of 117,513 tons or about 9.52% of the total national shallot production, followed by West Sumatra [2].

The productivity influences the productivity of NTB shallots from Bima Regency, designated by the government as a shallot development area. The planted area from 2014 is growing. From 2014 to 2016, they were 8,207, 10,491, and 13,674 ha, respectively, and the production was 91,709, 125,057 and 153,297 tons [2].

Shallots are mostly planted on marginal lands such as dry land, rainfed land, and under tree stands. The ground under tree stands usually has dry land conditions, so the plants get drought stress. Planting plants under tree stands causes plants to get shade stress, where plants experience a lack of sunlight. An understanding of shade stress is essential in onion planting. Shade stress affects all aspects of plant growth, including anatomical, morphological, physiological and biochemical plants. The yield of bulbs in shallots has been reported to be directly related to the amount of water supply and shade.

The primary constraint on the land under the stand is the low light intensity due to the shading factor. Therefore, efforts are needed to obtain adaptive and high-producing plant varieties with low light intensity (shade) conditions. Light has a significant role in physiological processes, such as photosynthesis, respiration, growth and development, closing and opening of stomata, various plant movements and germination [3].

The growth and morphology of shallots cannot be separated from environmental factors that influence them, one of which is shade. The presence of dye causes the intensity of sunlight plants

to receive to decrease. Plants will adapt, namely through changes in plant morphological characters. Changes in the morphology of these plants, such as the increase in plant height [4], decreased number of leaves and tillers [5], decreased leaf thickness [6] and increased leaf area [7, 8]. Environmental shade or low light intensity conditions also affect plant physiological conditions [9] and production [10, 11]. Authors [12] stated that physiological conditions include chlorophyll, carbohydrates, nitrogen levels, proteins, etc.

Plant tolerance determines the plants' response to the shaded environment to reduced light intensity. One of the effects of shade on plant morphology is that the plant stems become taller because the plant stems are etiolated [13]. This morphological condition causes the plant to fall, so it can quickly reduce tuber yields. According to [14, 15], shade changed the shallot bulb weight yield and influenced the growth process. Shade by 50 % during growth resulted in a decrease in soybean yield between 37 and 74 % [16, 17], and rice decreased yield productivity by more than 55 % [18].

Based on the description above, this study aimed to determine the physiological response and tolerance of several shallot genotypes to shade stress.

METHOD

The experimental design used was a completely randomised design (CRD) consisting of 6 genotypes, namely: G1 (Keta Monca), G2 (Ampanan), G3 (Super Philip), G4 (Thailand Nganjuk), G5 (Bali Karet), G6 (Lokananta). Each treatment was repeated five times. Planting was done using polybags measuring 35×35 cm; each polybag planted two bulbs, 2-3 cm deep, with a spacing of 20×20 cm. Shade treatment using 65.00% parent with a shade height of 2 m, a width of 1.5 m and a shade length of 3 m.

The implementation of the research includes providing seeds, preparing planting media, planting, maintaining, irrigating, fertilising, sampling, harvesting, post-harvesting and observing. There are seven parameters for keeping physiological responses: N-Total, protein, carbohydrates, carotenoids, chlorophyll a, chlorophyll b, and proline. N-Total analysis using the formula (1).

$$\text{Nitrogen content, \%} = \frac{(V_c - V_b) \times N \times 14}{\text{Sample weight}} \times 100, (1)$$

Protein analysis using the formula (2).

$$\text{protein content, \%} = N \times F_k$$

$$N = \frac{(HCl - \text{blank}) \times NHCL \times Fk \times 14}{\text{Sample weight}} \times 100, (2)$$

where V_c V_b – sample and blank titar, ml; N – normality of standard H_2SO_4 solution; 14 – equivalent weight of nitrogen; Fk = water content correction factor = $100/(100 - \% \text{ moisture content})$.

Carotenoid analysis using the formula (3).

$$\text{Carotenoids} = \frac{(A480 + 0.14A663 - 0.63A645) \times V \times Fp \times 10^3}{112.5W} (3)$$

where $A480$ – Absorbance at a wavelength of 480 nm; $A645$ – Absorbance at a wavelength of 645 nm; $A663$ – Absorbance at a wavelength of 663 nm; V – Extract volume, ml; W – Sample weight (g); Fp – Dilution factor.

Carbohydrate analysis using the formula (4).

$$\text{Carbohydrate, \%} = \frac{GT \times Fp}{\text{Sample weight}} \times 100, (4)$$

where GT – Table Sugar.

Chlorophyll analysis used the calculation formula (5, 6):

$$\text{Chlorophyll A} = 13.7A665 - 5.76A649 (5)$$

$$\text{Chlorophyll B} = 25.8A649 - 7.60A665 (6)$$

Proline analysis using the formula (7).

$$Y = 0.0336X + 0.0002 (7)$$

where Y – absorbance ; X – concentration.

Calculate the value of sensitivity (S) of shade stress based on the formula (8).

$$S = \frac{1 - \frac{Y}{Y_p}}{1 - \frac{X}{X_p}}, \quad (8)$$

where Y – score the mean of certain variables (e.g. several branches, plant height, and others) on one genotype under shade stress;

Y_p – score the average of these variables in an optimum environmental genotype;

X – score the average of these variables in all genotypes that experienced shade stress;

X_p – score the mean of these variables in all optimum environmental genotypes.

Score or Value: A – Shade stress tolerant genotype, $S < 0.5$; B – Somewhat tolerant, if $0.5 < S < 1$; C – Sensitive, if $S > 1$.

RESULTS AND DISCUSSION

Physiological Response of Several Genotypes of Shallots to Shade Stress. In Table 1, the analysis results show that the parameters of N-Total, protein, carbohydrates and carotenoids show significant differences in the genotype of shallots and indicate an interaction between treatments.

Table 1 – Physiological responses of several shallot genotypes on seven parameters of treatment

Parameter	Treatment		
	G	JN	G*JN
N-Total	S	S	S
Protein	S	S	S
Carbohydrate	S	S	S
Carotenoids	S	S	S
Chlorophyll a	NS	NS	NS
Chlorophyll b	NS	NS	NS
Proline	NS	NS	NS

Notes: G – Genotype; JN – Shade Type; G*JN – Combination of Genotype with Shade and No Shade Treatment; S – Significant; NS – Non-significant.

The parameters of chlorophyll a, chlorophyll b and proline showed no significant difference, and

there was no interaction between these treatments.

Each plant genotype has a different tolerance to shade stress. Plants adaptive to low radiation experience an increase in leaf area ratio, leaf stem ratio, and stem length and a decrease in leaf thickness [19]. Authors [20] reported that the interaction of the soybean genotype with the environment influenced the growth and yield of soybean plants. Research on the response of plant genotypes to shade is mostly carried out using modified shade environments such as a parent or black cloth. Artificial colouring is also widely used to analyse the growth and morphology of soybean plants [21, 22, 23].

In plants experiencing shade stress, plant height, according to [24], is more influenced by environmental factors, including light and water. Plant height showed etiolation symptoms characterised by excessive plant height growth. Authors [25] stated that plants with 50 % shade had higher growth but were etiolated. The hormone auxin influences this etiolation. In low light, auxin will spur higher stem growth, but the plant becomes weak, the stem is not sturdy, the leaves are small, and the plant looks pale. Symptoms of etiolation occur due to the absence or lack of sunlight. This is to the study's results, which showed an interaction between the shading treatment and the genotype of shallot plants on the parameters of N-Total, protein.

The results showed no interaction between the provision of shade and the genotype of shallot on the proline observation parameter. Proline compounds function to regulate cell osmotic pressure (osmotic adjustment). Accumulating proline can reduce the osmotic potential, reducing the water potential in cells without limiting enzyme function and maintaining cell turgor [26]. Authors [27] stated that there would be an increase in the proline content in the plant crown when the plant is under stress.

One of the critical external factors that can affect the growth and production of a plant is light intensity. Sunlight is one of the essential factors that can affect the photosynthesis rate in plants with chlorophyll. According to [28], sunlight has various effects on plant growth and provides a source of energy for photosynthesis. The absence of light will affect the physiological status of plant tissue so that the carbohydrate content will be reduced at low light intensity or dark. Changes in the level of endogenous hormones or other phys-

iological components can be affected by light intensity, duration, or quality of light. Light intensity is one of the critical factors for plant growth. Shade treatment can affect the chlorophyll content because the amount of light absorbed by plants is lower. The results showed that the chlorophyll observation parameter did not experience any interaction between the shading treatment and the onion genotype.

Table 2 – Average Value of Advanced Test Results (BNJ 5%) combination of shade and no shade treatments on the parameters of N-total, Protein, Carbohydrates and Carotenoids

Shade	Parameter			
	N-Total	Protein	Carbohydrate	Carotenoids
N ₀	0.22 ^a	1.35 ^a	4.63 ^b	104.59 ^b
N ₁	0.16 ^b	1.02 ^b	5.09 ^a	112.42 ^a
BNJ 5%	0.01	0.07	0.10	2.45
G1	0.17 ^{bc}	1.02 ^c	5.33 ^a	103.91 ^c
G2	0.23 ^a	1.43 ^a	4.32 ^d	114.36 ^a
G3	0.15 ^c	0.93 ^c	4.41 ^d	104.68 ^c
G4	0.22 ^a	1.37 ^a	4.78 ^c	101.65 ^c
G5	0.19 ^b	1.18 ^b	5.06 ^b	109.67 ^b
G6	0.19 ^b	1.17 ^b	5.29 ^a	116.76 ^a
BNJ 5%	0.02	0.12	0.18	4.25

Notes: Number-numbers followed by the same letter in the same column show no significant difference based on the BNJ test at a 5% significance level. G1= keta monca; G2= ampenan ; G3= super philip ; G4= tailland sulking; G5= rubber back; G6= lokananta.

The study's results (Table 1) showed that the effect of N-Total, protein, carbohydrates and carotenoids was significantly different on the genotype of shallot plants and indicated an interaction between the genotype treatment of shallot and the shade treatment. Shallots treated with no shade showed a better effect on N-Total and protein parameters than carbohydrates and carotenoids. The results showed that shading increased stem height, accelerated flowering time and reduced dry stover weight per plant. The increase in stem height is an attempt by plants to increase light absorption because plants cannot raise their leaves above the canopy. The research of [29] reported that shading caused an increase in stem height and leaf number of soybean plants.

The earlier flowering process occurs due to protein and carbohydrates (Table 1), which are readily soluble (phytochromes). Shaded environmental conditions can change the pigments

(phytochromes) in shaded soybean plants into forms that initiate flowering induction [30]. The decrease in the dry weight of plants due to shading is due to shady conditions where plants experience a limited amount of solar energy that can be absorbed for the optimal photosynthesis process, resulting in a decrease in the dry weight of the plants produced. As stated by [31], the dry weight is the net accumulation of carbon dioxide during plant growth.

The sunlight captured by chlorophyll raises the energy level of the electrons resulting from the oxidation of water in the process of photosynthesis. Plant energy can be used for biological purposes, macromolecular synthesis, and cell division. For example, division in the ontogenetic stomata system is based on how the guard stem cell divides, which is related to the direction of the cell wall that cuts that part of the meristemoïd [32].

Plants that grow in low light-intensity environments have smaller roots, are few and are composed of thin-walled cells. This occurs due to inhibiting the translocation of photosynthetic products from the roots. Longer plant stems are composed of thin-walled cells, larger intercellular spaces, and less transport and reinforcement tissue. The leaves are more significant and thinner, and the size of the stomata is more critical; the epidermal cells are light, but the number of leaves is less, and the space between cells is more. Experiments with iris leave grown at different intensities showed that the number of stomata decreased with decreasing light intensity. Responses to increased light intensity differ between plants suitable for shaded conditions (shade plants; indoor plants); with plants that can grow in unshaded conditions. Shade-suitable plants show meagre photosynthetic rates at high light intensities [33]. The photosynthetic rate of shaded plants reaches saturation point at lower light intensities, higher photosynthetic rates at very low light intensities, and lower light compensation points than open-fit plants. From the above description, plants suitable for shade can survive in shaded conditions (low light intensity), while plants suitable for open cannot survive [33].

Tolerance of several Shallot Genotypes to Shade Stress. Tolerance is a mechanism for plant resistance in a state of shade stress and affects production, reducing crop yields or productivity. Based on the formula of [34], the genotype of

shallots tolerant of shade stress was obtained. Table 3 shows that the sensitive plant genotypes (P) to shade stress are G3, G4, and G5. The moderately tolerant genotype (AT) was G2, and the tolerant genotype (T) was G1.

Table 3 – Value of Sensitivity Index (S) of six Shallot Genotypes tested under Shade Stress and No Shade Stress

Genotype	Sensitivity Value (S)		Average	Phenotype
	BKU	JU		
G1	0.21	0.29	0.25	T
G2	1.35	0.48	0.92	AT
G3	0.15	3.31	1.73	P
G4	1.40	0.60	1.00	P
G5	2.33	0.34	1.34	P
G6	0.61	1.04	0.83	AT

Notes :S=Sensitivity Index Value; T= Tolerant (< 0.5); AT= Somewhat Tolerant (>0.5 $S < 1$); P=Sensitive ($S > 1$); BKU= Bulb Dry Weight; JU= Number of Bulbs. G1= keta monca; G2= ampenan ; G3= super philip ; G4= tailland sulking; G5= rubber back; G6=lokananta.

From Table 3, the G1 genotype (Keta Monca) shows a genotype that is tolerant to shade stress. Keta Monca is a local variety of shallots that develops mainly in the Bima Regency, West Nusa Tenggara Province. Production is relatively suitable for local types, has a distinctively spicy taste, and is highly flavourful and savoury. This shallot has a pretty good market, especially in big cities in Indonesia. The shallot cultivation business in Indonesia uses two sources of planting material: local and imported. Farmers prefer to use local varieties because the age of harvest is in line with expectations, and by the conditions of the agroecosystem, the ease of farmers in managing, and plant care that is by the habits of farmers.

In onion plants, shade is leaf protection from damage by exposure to rainwater. This Keta Monca genotype is also a plant resistant to Fusarium wilt disease [35]. When watering, it is adjusted to plant conditions (field capacity) and the weather at the planting time because shallot plants are susceptible to disease if the soil is too wet or too dry. The regulation of osmotic pressure in plants is the work of several compounds that act as osmo-regulators, one of which is proline. Proline Accumulation can reduce osmotic

potential, thereby reducing water potential in cells without limiting enzyme function and maintaining cell turgor [36].

Shallots require full light intensity with irradiation of more than 12 hours [37] and air temperatures between 20–30 °C with an optimal average temperature of 24 °C [38].

Sunlight is also thought to affect the growth of shallot plants. Shade conditions prevent sunlight from entering the research site. This results in a lack of sunlight received by plants, thereby inhibiting the process of photosynthesis for plant growth. According to [39], sunlight is an essential factor for the ongoing process of photosynthesis, while photosynthesis is very important for plant metabolism. Added to [40], the effect of sunlight on plants is focused on vegetative and generative growth. Plant response to light is determined by green leaf synthesis, stomata activity (respiration, transpiration), anthocyanin formation, temperature from surface organs, absorption of mineral nutrients, permeability, respiratory rate, and protoplasm flow. Theoretically, the greater the amount of available energy will increase photosynthesis. Lack of sunlight will interfere with photosynthesis and growth, although the need for light depends on the type of plant itself.

Authors [41] stated that the level of shading treatment had no significant effect on the production of fresh matter and dry matter production of forage. Authors [42] noted that the performance of mung bean yields in the 75 % shade reduced the output of mung bean to 65.21% when compared to no shade, but when compared to 50 % shade, the yield of mung bean only decreased by 34.01%.

The results of [43] also showed that shading stress applied from the beginning of the plant to harvest resulted in long stem segments of peanut plants. Authors [44] reported that the intensity of shading up to 75 % increased plant height and reduced the number of leaves on soybean plants.

In addition, lack of light during plant development will cause etiolation symptoms, where plant stems will grow faster but weaker, and the leaves are small, thin and pale (not green). A lack of light causes symptoms of etiolation, or the plant is in a dark and shaded place. Light can also act as an inhibitor in the growth process. This is because it can stimulate the diffusion of auxin to plants not exposed to sunlight. The light that is an inhibitor is caused by the absence of light so that

it can maximise the function of auxin to support plant cells.

On the contrary, plants that grow in bright places cause plants to grow more slowly with relatively short, more comprehensive, greener conditions, look fresher, and the plant stems are stronger because sunlight is essential and influences plant growth and development [45].

CONCLUSIONS

The results of the physiological response research showed that the parameters of N-Total, protein, carbohydrates and carotenoids showed significant differences in the genotype of shallots and indicated an interaction between treatments. The parameters of chlorophyll a, chlorophyll b and proline showed no significant difference and no interaction between treatments. The tolerant shallot genotypes (T) were G1 (Keta Monca), and the slightly tolerant genotypes (AT) were G2 (Ampanan) and G6 (Lokananta).

REFERENCES

1. Direktorat Bina Produksi Hortikultura Provinsi Sumatera Barat. (1999). *Laporan Tahunan Dinas Tanaman Pangan dan Hortikultura Sumatera Barat* [Annual Report of the Food Crops and Horticulture Service of West Sumatra]. Padang (in Indonesian).
2. Sekretaris Direktorat Jenderal Hortikultura. (2015). Statistik Produksi Hortikultura Tahun 2014 [Horticulture Production Statistics 2014]. Retrieved from <https://pdfslide.tips/documents/statistik-produksi-2014.html?page=1> (in Indonesian).
3. Taiz, L., & Zeiger, E. (2003). *Plant Physiology*. California: The Benjamin Pub.
4. Thakur, P. K., Nikam, B. R., Srivastav, S. K., Wint Khaing, T. W., Zaw, T., Garg, V., Dhote, P. R., Sharma, V., & Aggarwal, S. P. (2021). Groundwater modeling with inputs from geospatial technology for assessing the sustainability of water use in the Solani watershed, Ganga river basin (India). *Groundwater for Sustainable Development*, 12, 100511. doi: [10.1016/j.gsd.2020.100511](https://doi.org/10.1016/j.gsd.2020.100511)
5. Buntoro, B. H., Rogomulyo, R., & Trisnowati, S. (2014). *Pengaruh takaran pupuk kandang dan intensitas cahaya terhadap pertumbuhan dan hasil temu putih (Curcuma zedoaria L.)* [Effect of manure dose and light intensity on growth and yield of white temu (*Curcuma zedoaria* L.)]. *Vegetalika*, 3(4), 29–39 (in Indonesian).
6. Fan, Y., Chen, J., Cheng, Y., Raza, M. A., Wu, X., Wang, Z., Liu, Q., Wang, R., Wang, X., Yong, T., Liu, W., Liu, J., Du, J., Shu, K., Yang, W., & Yang, F. (2018). Effect of shading and light recovery on the growth, leaf structure, and photosynthetic performance of soybean in a maize-soybean relay-strip intercropping system. *PLOS ONE*, 13(5), e0198159. doi: [10.1371/journal.pone.0198159](https://doi.org/10.1371/journal.pone.0198159)
7. Umarie, I., Widarti, W., Wijaya, I., & Hasbi, H. (2018). *Pengaruh warna naungan plastik dan dosis pupuk organik kompos terhadap pertumbuhan bawang merah (Allium ascalonicum L.)* [Effect of plastic shade colour and compost organic fertiliser dosage on shallot growth (*Allium ascalonicum* L.)]. *Jurnal Agroqua*, 16(2), 129–137 (in Indonesian).
8. Ritonga, A. W., Chozin, M. A., Syukur, M., Maharijaya, A., & Sobir. (2019). Heritabilitas, Korelasi, dan Sidik Lintas Berbagai Karakter Tomat pada Kondisi Naungan dan Tanpa Naungan [Heritability, Correlation, and Cross-Sections of Various Tomato Characters under Shade and Shade-free Conditions]. *Jurnal Hortikultura Indonesia*, 10(2), 85–93. doi: [10.29244/jhi.10.2.85-93](https://doi.org/10.29244/jhi.10.2.85-93) (in Indonesian).
9. Wulandari, R., Suminarti, N., & Sebayang, H. (2016). *Pengaruh jarak tanam dan frekuensi penyiangan gulma pada pertumbuhan dan hasil tanaman bawang merah (Allium ascalonicum)* [Effect of spacing and frequency of weeding on the growth and yield of shallot (*Allium ascalonicum*) plants]. *Jurnal Produksi Tanaman*, 4(7), 547–553 (in Indonesian).
10. Baharuddin, R., Chozin, M., & Syukur, M. (2014). *Toleransi 20 Genotipe tanaman tomat terhadap naungan* [Toleransi 20 Genotipe Tanaman Tomat terhadap Naungan]. *Indonesian Journal of Agronomy*, 42(2) (in Indonesian).

11. Wakhid, R., Widodo, S., Pudjojono, M. (2012). Pengaruh pemberian naungan dan mulsa terhadap kadar air tanah dalam produksi tanaman bawang merah pada musim penghujan [Effect of shading and mulching on soil moisture content in shallot crop production in the rainy season]. *Jurnal Agroteknologi*, 6(1), 51–58 (in Indonesian).
12. Ulinnuha, Z., Ahmad Chozin, M., & Santosa, E. (2019). Stabilitas Hasil dan Gangguan Penyakit pada Enam Genotipe Tomat di bawah Naungan [Yield Stability and Disease Disturbance in Six Tomato Genotypes under Shade]. *Jurnal Hortikultura Indonesia*, 10(1), 10–19. doi: 10.29244/jhi.10.1.10-19 (in Indonesian).
13. Uchimiya, H. (2001). *Genetic engineering for abiotic stress tolerance in plants*. Retrieved from <https://agris.fao.org/agris-search/search.do?recordID=XF2016018151>
14. Tambunan, W., Sipayung, R., Sitepu, F. (2014). Pertumbuhan Dan Produksi Bawang Merah (*Allium Ascalonicum* L.) Dengan Pemberian Pupuk Hayati Pada Berbagai Media Tanam [Growth and Production of Red Onion (*Allium Ascalonicum* L.) with Biofertilisation in Various Planting Media]. *Jurnal Online Agroekoteknologi*, 2(2), 825–836 (in Indonesian).
15. Sumarni, N., & Rosliani, R. (2010). Pengaruh Kerapatan Tanaman, Naungan, dan Mulsa Terhadap Pertumbuhan dan Produksi Umbi Bawang Merah Mini Asal Biji [Effect of Plant Density, Shade, and Mulch on Growth and Production of Seed-origin Mini Shallot Bulbs]. *Jurnal Hortikultura*, 12(1), 28–34 (in Indonesian).
16. Chotechuen, S. (1996). Breeding of mungbean for resistance to various environmental stresses. In C. Kitbamroong, S. Miyazaki (Eds.), *Mungbean Germplasm: Collection, Evaluation and Utilization for Breeding Program* (pp. 52–59). Bangkok: Proceedings of the Workshop on Mungbean Germplasm.
17. Steppuhn, H., van Genuchten, M. Th., & Grieve, C. M. (2005). Root-Zone Salinity. *Crop Science*, 45(1), doi: 10.2135/cropsci2005.0209
18. Sulistyono, E., Chozin, M., & Rezkiyanti, F. (2002). Uji potensi hasil beberapa galur padi gogo (*Oryza sativa* L.) pada beberapa tingkat naungan [Test of yield potential of some upland rice (*Oryza sativa* L.) strains in several shade levels]. *Jurnal Agronomi Indonesia*, 30, 1–5 (in Indonesian).
19. Haque, M. M., Hasanuzzaman, M., & Rahman, M. (2009). Effect of light intensity on the morpho-physiology and yield of bottle gourd (*Lagenaria vulgaris*). *Academic Journal of Plant Sciences*, 2(3), 158–161.
20. Sundari, T., Nugrahaeni, N., & Susanto, G. (2016). Interaksi genotipe x lingkungan dan stabilitas hasil biji kedelai toleran naungan [Genotype x Environment Interaction and Stability of Grain Yield of Shade Tolerant Soybean]. *Jurnal Agronomi Indonesia*, 44, 16–25 (in Indonesian).
21. Muhuria, L., Tyas, K., Khumaida, N., Trikoessoemaningtyas, & Sopandie, D. (2006). Adaptasi kedelai terhadap intensitas cahaya rendah: Karakter daun untuk efisiensi penangkapan cahaya [Soybean Adaptation to Low Light Intensity: Leaf Characters for the Light Capture Efficiency]. *Jurnal Agronomi Indonesia*, 34(3), 133–140 (in Indonesian).
22. Kisman, Khumaida, N., Trikoessoemaningtyas, Sopandie, D. (2007). Karakter morfo-fisiologi daun, penciri adaptasi kedelai terhadap intensitas cahaya rendah [Leaf morpho-physiological characters, characterising soybean adaptation to low light intensity]. *Jurnal Agronomi Indonesia*, 35(2), 96–102 (in Indonesian).
23. Chairudin, Efendi, & Sabaruddin. (2015). Dampak naungan terhadap perubahan karakter agronomi dan morfofisiologi daun pada tanaman kedelai (*Glycine max* (L.) Merrill) [Impact of Shade to Changes of Characters of Agronomy and Morpho-Physiology Leaves in Soybean (*Glycine Max* (L.) Merrill)]. *Jurnal Floratek*, 10, 26–35 (in Indonesian).
24. Sitompul, S. (2016). *Analisis Pertumbuhan Tanaman* [Plant Growth Analysis]. Malang: UB Press (in Indonesian).

25. Tanari, Y., & Vita, V. (2017). Pengaruh naungan dan berbagai media tanam terhadap pertumbuhan dan produksi tanaman selada (*Lactuca sativa* L.) [Effect of shade and various growing media on the growth and production of lettuce (*Lactuca sativa* L.) plants]. *Jurnal AgroPet*, 14(2) (in Indonesian).
26. Tuasamu, Y. (2009). *Toleransi hotong (Setaria italica l. Beauv) pada berbagai cekaman kekeringan: pendekatan anatomi dan fisiologi* [Tolerance of hotong (*Setaria italica* l. Beauv) to various drought stresses: anatomical and physiological approaches] (Master's thesis); Institut Pertanian Bogor. Retrieved from <https://repository.ipb.ac.id/handle/123456789/5960> (in Indonesian).
27. Maestri, M., Da Matta, F. M., Regazzi, A. J., & Barros, R. S. (1995). Accumulation of proline and quaternary ammonium compounds in mature leaves of water stressed coffee plants (*Coffea arabica* and *C. canephora*). *Journal of Horticultural Science*, 70(2), 229–233. doi: 10.1080/14620316.1995.11515292
28. Yuliarti, N. (2010). *Kultur Jaringan Skala Rumah Tangga* [Household Scale Tissue Culture]. Yogyakarta: Andi (in Indonesian).
29. Bakhshy, J., Golezani, K., Zehtab-Salmasi, S. & Moghaddam, M. (2013). Effects of water deficit and shading on morphology and grain yield of soybean (*Glycine max* L.). *Technical Journal of Engineering and Applied Sciences*, 3(1), 39–43.
30. Karamoy, L. (2009). Hubungan iklim dengan pertumbuhan kedelai (*Glycine max* (L.) Merrill) [Climate relationship with soybean (*Glycine max* (L.) Merrill) growth]. *Soil Environment*, 7, 65–68 (in Indonesian).
31. Gardner, F. (1991). *Fisiologi Tanaman Budidaya* [Plant Physiology of Cultivation]. Jakarta: Universitas Indonesia Press (in Indonesian).
32. Fahn, A. (1992). *Anatomi Tumbuhan* [Plant Anatomy]. Jakarta: Gramedia (in Indonesian).
33. Lakitan. (1993). *Dasar-dasar Fisiologi Tumbuhan* [Fundamentals of Plant Physiology]. Jakarta: Raja Grafindo Persada (in Indonesian).
34. Fischer, R., & Maurer, R. (1978). Drought resistance in spring wheat cultivars. I. Grain yield responses. *Australian Journal of Agricultural Research*, 29(5), 897. doi: 10.1071/ar9780897
35. BALISTA. (2011). *Bawang merah varietas bima brebes* [Onion variety bima brebes]. Retrieved from <http://balitsa.litbang.pertanian.go.id/ind/index.php/berita-terbaru/1132-bawang-merah-varietas-bima-brebes> (in Indonesian).
36. Kurniawati, S., Khumaida, N., Ardie, S., Hartati, N., Sudarmonowati, E. (2014). Pola Akumulasi Prolin dan Poliamin Beberapa Aksesori Tanaman Terung pada Cekaman Kekeringan [Proline and Polyamines Accumulation Pattern of Some Eggplant Accessions under Drought Stress]. *Jurnal Agronomi Indonesia*, 34(3), 133–140 (in Indonesian).
37. Currah, L., Proctor, F. (1990). *Onion in Tropical Region*. *National Resources Institute Bulletin*, 35, 20–21.
38. Grubben, G. J. H. (1990). Timing of Vegetable Production In Indonesia. *Acta Horticulturae*, 267, 261–270. doi: 10.17660/actahortic.1990.267.33 (in Indonesian).
39. Kramer, P., Kozlowsky, L. (1979). *Physiology of Woods Plants*. Florida: Academic Press.
40. Jumin, H. (2008). *Dasar-Dasar Agronomi* [Fundamentals of Agronomy]. Jakarta: Rajagrafindo Persada (in Indonesian).
41. Hanafi, D., Umar, S., & Bachri, I. (2005). Pengaruh tingkat naungan pada berbagai pastura campuran terhadap produksi hijauan [Effect of shade levels in various mixed pastures on forage production]. *Jurnal Agibisnis Peternakan*, 1(3), 101–105 (in Indonesian).
42. Sundari, T., Soemartono, Tahari, Mangoendidjojo, W. (2005). Keragaan hasil dan toleransi genotipe kacang hijau terhadap penanungan [Yield performance and tolerance of mung bean genotypes to damping off]. *Jurnal ilmu pertanian*, 12(1), 12–19 (in Indonesian).

43. Hemon, F. (2020). Pengaruh intensitas cahaya rendah pada berbagai fase pertumbuhan terhadap karakter morfologi dan daya hasil kacang tanah [Effect of low light intensity at different growth stages on morphological characters and yield of groundnut]. Seminar Nasional Peragi (in Indonesian).
44. Sundari, T., & Anggoro Susanto, G. W. (2016). Pertumbuhan dan Hasil Biji Genotipe Kedelai di Berbagai Intensitas Naungan [Growth and Seed Yield of Soybean Genotypes at Different Shade Intensities]. *Jurnal Penelitian Pertanian Tanaman Pangan*, 34(3), 203. doi: [10.21082/jpntp.v34n3.2015.p203-217](https://doi.org/10.21082/jpntp.v34n3.2015.p203-217) (in Indonesian).
45. Fitter, A., Hay, R. (1991). *Fisiologi Lingkungan Tanaman* [Plant Environmental Physiology]. Yogyakarta: Gadjah Mada University Press (in Indonesian).